

CD146 acts as a novel receptor for netrin-1 in promoting angiogenesis and vascular development

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Angiogenesis, a process that newly-formed blood vessels sprout from pre-existing ones, is vital for vertebrate development and adult homeostasis. Previous studies have demonstrated that the neuronal guidance molecule netrin-1 participates in angiogenesis and morphogenesis of the vascular system. Netrin-1 exhibits dual activities in angiogenesis: either promoting or inhibiting angiogenesis. The anti-angiogenic activity of netrin-1 is mediated by UNC5B receptor. However, how netrin-1 promotes angiogenesis remained unclear. Here we report that CD146, an endothelial transmembrane protein of the immunoglobulin superfamily, is a receptor for netrin-1. Netrin-1 binds to CD146 with high affinity, inducing endothelial cell activation and downstream signaling in a CD146-dependent manner. Conditional knockout of the *cd146* gene in the murine endothelium or disruption of netrin-CD146 interaction by a specific anti-CD146 antibody blocks or reduces netrin-1-induced angiogenesis. In zebrafish embryos, downregulating either netrin-1a or CD146 results in vascular defects with striking similarity. Moreover, knocking down CD146 blocks ectopic vascular sprouting induced by netrin-1 overexpression. Together, our data uncover CD146 as a previously unknown receptor for netrin-1 and also reveal a functional ligand for CD146 in angiogenesis, demonstrating the involvement of netrin-CD146 signaling in angiogenesis during vertebrate development.

Keywords: angiogenesis; CD146; netrin-1; vascular development

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Introduction

Angiogenesis, a process in which newly-formed blood vessels sprout from pre-existing ones, is vital for development and adult homeostasis [1]. Angiogenesis is also an important aspect in a large number of human diseases [2]. During angiogenesis, endothelial cells degrade the basement membrane, migrate into the extracellular space, proliferate and form capillary sprouts and tubular structures. All these processes are tightly regulated by a network of pro- and anti-angiogenic factors [3, 4],

among which the vascular endothelial growth factor (VEGF) and its functional endothelial receptor VEGFR2 are major contributors [5, 6]. Recent studies have established that the four major classes of neuronal guidance molecules, including netrins, Slits, semaphorins and ephrins, play important roles in angiogenesis and pattern formation during vascular morphogenesis [7-12]. Netrin family members are secreted proteins that promote axon outgrowth and guide growth cone navigation during neural development [13]. They share an N-terminal type VI laminin repeat (domain VI), followed by three cysteine-rich epidermal growth factor modules (domains V) and a positively charged C-terminus (netrin-like domain, NTR). Netrin-1, a prototype of netrin family, can act to either attract or repel growing axons during neural development [14, 15]. The netrin-1 receptor Deleted in Colon Cancer (DCC) can mediate both attractive and repul-

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sive responses [16, 17], whereas members of the UNC5 family mediate netrin-1-induced repulsion [18, 19]. In angiogenesis, netrin-1 also has dual functions: either promoting or inhibiting endothelial cell activation and angiogenesis [7, 8, 20, 21]. UNC5B, the only known cognate receptor for netrin-1 on endothelial cells, mediates the anti-angiogenic effect of netrin-1 [7, 21]. However, the molecular mechanisms by which netrin-1 promotes angiogenesis remain to be elucidated.

CD146 (also known as melanoma cell adhesion molecule, MCAM) is a member of cell adhesion molecules of the immunoglobulin (Ig) superfamily [22]. It contains five Ig-like domains in its extracellular region, a single transmembrane domain and a cytoplasmic tail [23]. As a multi-functional molecule, CD146 participates in various biological processes including angiogenesis, tumor metastasis, lymphocyte activation, morphogenesis during development and tissue regeneration [24–26]. Expressed on endothelial cells, CD146 is required for endothelial cell proliferation, migration and tube formation [27, 28], playing critical roles in angiogenesis [29–32]. In our previous studies, we have developed a CD146-specific monoclonal antibody (mAb) AA98 that blocks tumor angiogenesis both *in vitro* and *in vivo* [29, 33]. In addition, CD146 serves as a co-receptor for VEGFR2 to facilitate the transduction of VEGF signaling in endothelial cells [34]. In spite of intensive studies, specific ligand(s) involved in CD146-mediated angiogenesis has not been identified.

In this study, we report that netrin-1 binds to CD146 with high affinity and the netrin-CD146 interaction is required for netrin-1-induced endothelial cell activation, as well as downstream VEGF signal transduction. In mouse, deletion of endothelial CD146 or disruption of netrin-CD146 interaction by anti-CD146 mAb AA98 impairs netrin-1-induced angiogenesis. In zebrafish embryos, downregulation of netrin-1a or CD146 results in strikingly similar vascular defects. Importantly, knocking down CD146 blocks ectopic vascular sprouting and branching induced by netrin-1 overexpression. Our findings demonstrate that CD146 is a previously unknown receptor required for netrin-1-induced angiogenesis and support a key role of netrin-CD146 signaling in angiogenesis during vertebrate development.

Results

Netrin-1 interacts with CD146

In an effort to identify new ligands for CD146 in angiogenesis, we tested netrin-1 for its potential interaction with CD146 using biochemical and cell biological approaches. In Human Embryonic Kidney 293 (HEK293)

cells that co-transfected with netrin-1- and CD146-expressing plasmids, both netrin-1 and CD146 could be immunoprecipitated by either anti-CD146 or anti-netrin-1 antibody but not by the mouse IgG (mIgG) control (Figure 1A), indicating that netrin-1 interacts with CD146.

To test whether netrin-CD146 interaction could occur on the surface of living cells, we employed a cell surface binding assay. Following transfection of HEK293 cells with plasmids encoding DCC receptor (as a positive control), CD146 or Robo-1 (as a negative control), cells were incubated with conditional media prepared from netrin-1-GFP stable expression cells. Fluorescent microscopy was used to detect netrin-1-GFP signals on the cell surface. Netrin-1-GFP binding signals were detected on HEK293 cells that expressing either DCC or CD146, but not Robo-1 (Figure 1B). Robo-1 is another member of the Ig superfamily that expressed on growing axons and serves as a receptor for Slits [35]. These results support a specific interaction between netrin-1 and CD146.

To demonstrate a direct interaction between netrin-1 and CD146, we performed a pull-down assay using purified proteins. The Fc fragment of human IgG (Fc, as a control), the extracellular domain of CD146 or UNC5B fused to the Fc fragment (Fc-CD146 or Fc-UNC5B) was first bound to protein G sepharose beads and then incubated with purified netrin-1 protein. Netrin-1 interacted with Fc-CD146 or Fc-UNC5B, whereas Fc alone did not show binding activity (Figure 1C), indicating that the extracellular region of CD146 directly binds to netrin-1. Furthermore, the binding affinity of netrin-1 to CD146 or UNC5B was measured using surface plasmon resonance (SPR). Interestingly, the apparent dissociation constant (K_d) of netrin-CD146 interaction was ~ 1.33 nM (Figure 1D), lower than that of netrin-UNC5B interaction with a K_d value of 5.10 nM (Figure 1E).

We next mapped the domains required for netrin-CD146 interaction using netrin-1 and CD146 truncation mutant constructs. In the co-immunoprecipitation experiments, CD146 was immunoprecipitated by the full-length netrin-1 or its truncation mutant Δ NTR, in which the C-terminal NTR domain of netrin-1 was deleted. However, further deletion to remove the domain V (Δ NTR-V) of netrin-1 abolished the interaction between netrin-1 and CD146 (Figure 2A and 2B). These results indicate that the domain V of netrin-1 is critical for its binding to CD146. Similarly, when using a series of CD146 truncation mutant constructs with the extracellular Ig domains deleted sequentially, netrin-1 was immunoprecipitated by the full-length CD146, or the truncation mutants, Δ D1, Δ D1-D2 and Δ D1-D3. However, further deletion to remove the 4th Ig domain (Δ D1-D4) abolished the interaction (Figure 2C and 2D), indicating

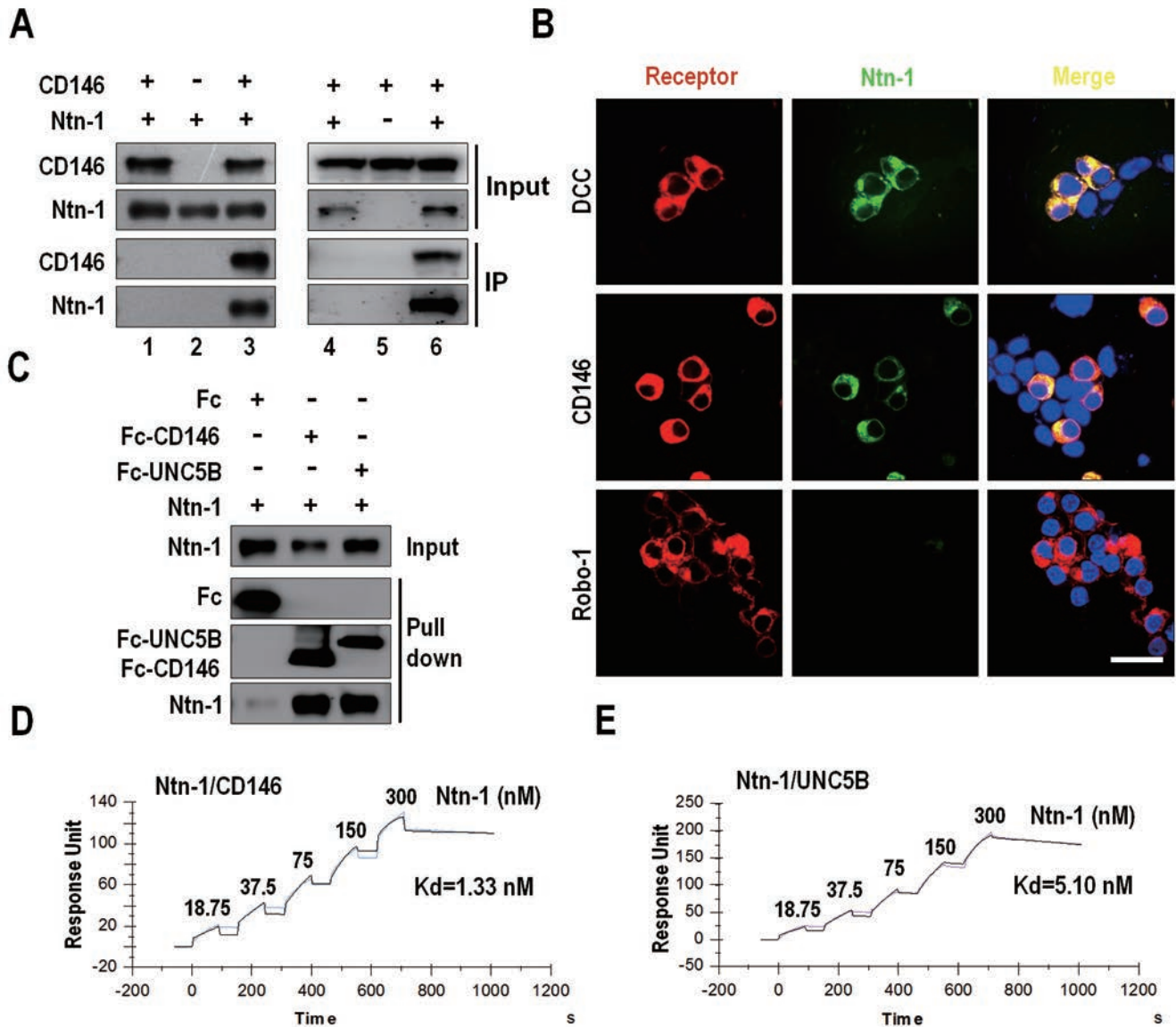


Figure 1 Netrin-1 binds to CD146. **(A)** Co-immunoprecipitation assays. Netrin-1- and CD146-expressing plasmids were co-transfected into HEK293 cells prior to preparation of cell lysates. Lanes 1 and 4: precipitated by control mIgG. Lanes 2 and 3: anti-CD146 mAb AA1. Lanes 5 and 6: anti-netrin-1 mAb. **(B)** HEK293 cells were transfected with plasmids encoding DCC, CD146 or Robo-1 and incubated with netrin-1-GFP conditional medium. Binding of netrin-1-GFP to the cell was detected by its GFP fluorescence. Scale bar, 50 μ m. **(C)** Direct interaction between purified netrin-1 and CD146 proteins *in vitro*. Fc, Fc-CD146 or Fc-UNC5B (200 ng/ml) bound protein G beads was incubated with netrin-1 protein (200 ng/ml). The bound proteins were analyzed by western blotting. **(D, E)** Determination of netrin-CD146 binding affinity by SPR. Purified netrin-1 protein was applied at different concentrations to the CM5 chips containing Fc-CD146 **(D)** or Fc-UNC5B **(E)**. Data represent 3 independent experiments.

the 4th Ig domain of CD146 is critical for the binding to netrin-1.

In our previous studies, we have developed a specific function-blocking mAb against CD146, AA98, which recognizes CD146 at its 4th-5th Ig domains [29, 36]. As the 4th Ig domain of CD146 is required for netrin-CD146

interaction, we tested whether AA98 could block the binding. In the pull-down assay in which two anti-CD146 mAbs, AA1 and AA98, were first bound to protein G beads and then incubated with CD146 extracellular region (soluble CD146 (sCD146)) and netrin-1, netrin-1 could be precipitated by AA1 but not AA98 (Supplemen-

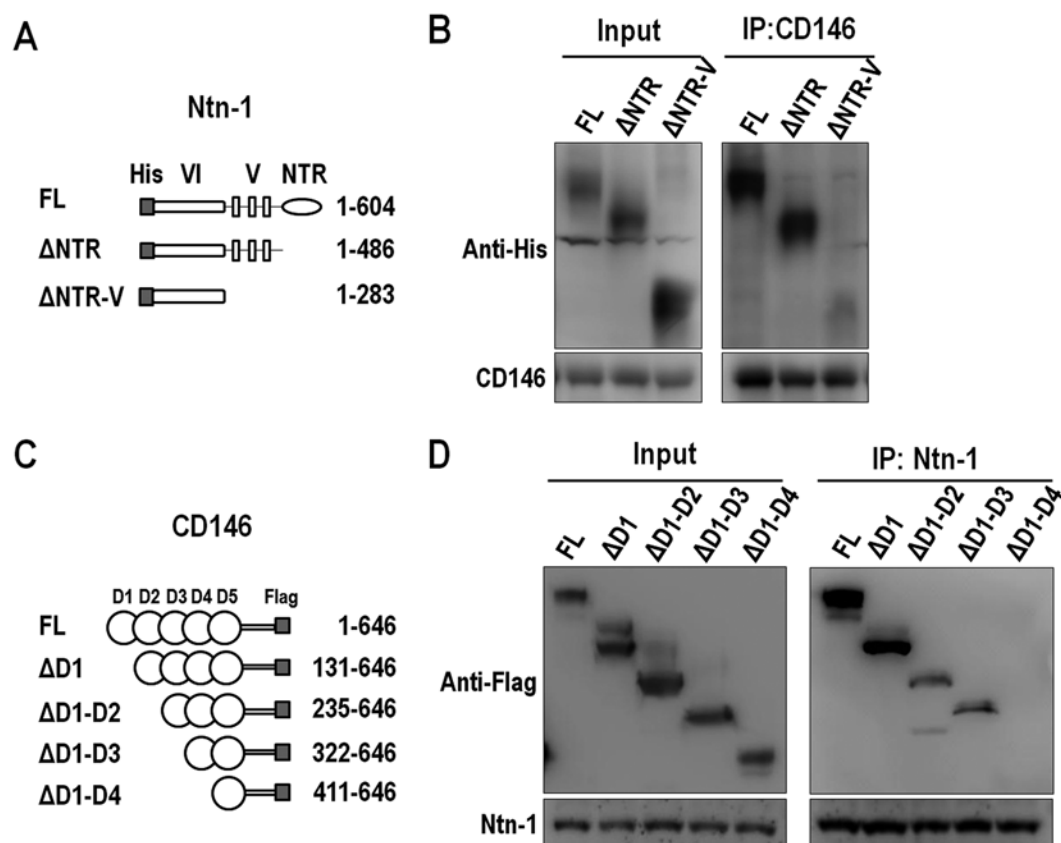


Figure 2 Identification of interacting domains of netrin-1 and CD146. **(A)** A diagrammatic illustration of the full-length (FL) and truncation mutants of netrin-1 expressed as His-tagged proteins. **(B)** HEK293 cells were co-transfected with plasmid expressing CD146 and the FL or truncation mutant of netrin-1. The cell lysates were prepared for immunoprecipitation with anti-CD146 mAb AA1. **(C, D)** Defining the domain of CD146 required for interaction with netrin-1. CD146 and its truncation mutants were expressed as Flag-tagged proteins. Immunoprecipitation was performed using anti-netrin-1 mAb. Data represent 3 independent experiments.

tary information, Figure S1A). AA1 recognizes CD146 at its 1st-2nd Ig domains of the extracellular region [36]. Similar results were obtained in a cell surface binding assay. When the HEK293 cells were pre-treated with mAb AA98, the binding of netrin-1 to CD146-expressing cells was inhibited (Supplementary information, Figure S1B). These results support the notion that AA98 was capable of blocking netrin-CD146 interaction. Thus, AA98 was used as a blocker for netrin-CD146 interaction in subsequent functional assays.

CD146 is required for netrin-1-induced endothelial cell activation

We examined the effect of netrin-1 on endothelial cell activation using human umbilical vein endothelial cells (HUVECs). Low concentrations of netrin-1 (50 or 200 ng/ml) promoted HUVEC proliferation (Figure 3A), migration (Figure 3B and 3E) and tube formation (Figure

3C and 3E). In contrast, netrin-1 at 2 000 ng/ml exhibited inhibitory effect. These results are in line with previous studies [37]. To delineate downstream pathways mediating netrin-1-induced endothelial cell activation, we first examined the dimerization of CD146, an important mechanism required for its downstream signaling [28]. In HUVECs, netrin-1 induced CD146 dimerization in a dose-dependent manner (Figure 3D). Interestingly, in a recent study, Xu *et al.* [38] reported that the molecular structure of netrin-1 contains two receptor-binding ends to bring together receptor molecules. Because CD146 could serve as a co-receptor for VEGFR2 and facilitate the VEGF signal transduction [34], we next tested whether VEGF signaling is activated by netrin-1 treatment. Consistent with the cell functional assays, netrin-1 at 50 or 200 ng/ml induced the phosphorylation of VEGFR2, ERK1/2 and p38, whereas at 2 000 ng/ml it inhibited the phosphorylation of these proteins (Figure 3D). ERK1/2

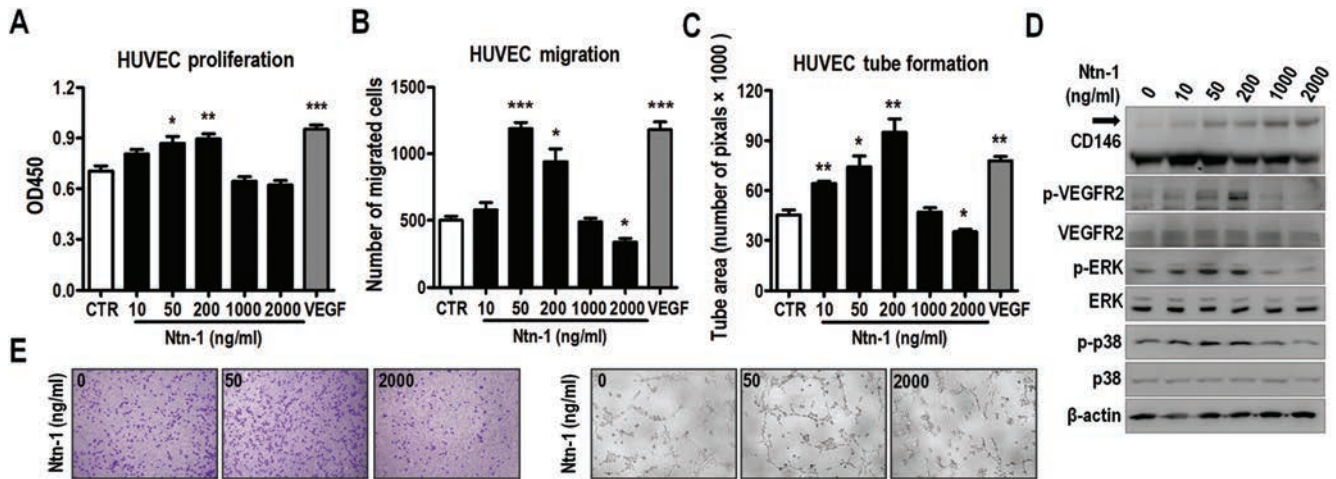


Figure 3 Netrin-1 exhibits dual activities in endothelial cell activation. **(A–C)** HUVECs were used for proliferation assay **(A)**, transwell migration assay **(B)** and tube formation assay **(C)**. Netrin-1 was applied at concentrations as indicated. **(D)** serum-starved HUVECs were treated with netrin-1 at different concentrations for 10 min. Dimerization of CD146 and phosphorylation of VEGFR2, ERK1/2 and p38 were analyzed by western blotting. The black arrow marks the CD146 dimer. **(E)** Representative images of netrin-1-induced HUVEC migration (left) and tube formation (right). For **A–C**, $n = 3$ in each group. Data represent 3 independent experiments (means \pm SEM). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$ (one-way ANOVA with Turkey *post hoc* tests).

and p38 are key signaling molecules downstream of VEGFR2 that contribute to endothelial cell activation and angiogenesis [5]. Netrin-1 did not bind to VEGFR2, as shown by the co-immunoprecipitation (Supplementary information, Figure S2A) and pull-down assays (Supplementary information, Figure S2B), suggesting that netrin-1 affects VEGF signaling through its interaction with CD146.

Previous studies have demonstrated that the inhibitory effect of netrin-1 on angiogenesis depends on UNC5B [7, 21], the only known cognate receptor of netrin-1 that expressed on endothelial cells. Here we investigated the roles of CD146, UNC5B and VEGFR2 in mediating the dual activities of netrin-1. Efficient downregulation of CD146 expression by its specific siRNA abolished netrin-1 (at 50 or 200 ng/ml) induced HUVEC proliferation (Figure 4A), migration (Figure 4B) and tube formation (Figure 4C), as well as VEGF signaling activation (Figure 4D), whereas the inhibitory effect of netrin-1 at 2 000 ng/ml were not affected. In contrast, downregulation of UNC5B receptor did not affect the pro-angiogenic activities of netrin-1 at low concentrations, but converted the inhibitory effect of netrin-1 at 2 000 ng/ml to pro-angiogenic effect (Figure 4), suggesting that in the absence of UNC5B receptor, netrin-1 mainly promotes endothelial cell activation via CD146. These results indicate that CD146 and UNC5B mediate the pro- and anti-angiogenic activities of netrin-1, respectively. Moreover, knocking

down VEGFR2 partially reduced but did not abolish netrin-1-induced HUVEC activation (Figure 4A–4C), suggesting that other pathways may also be involved downstream of netrin-CD146 to contribute to endothelial cell activation. In contrast, downregulation of CD146 abolished VEGF-induced HUVEC migration and tube formation but not proliferation (Figure 4A–4C), indicating a critical role of CD146 in mediating VEGF signaling, which is consistent with our previous study [34].

To confirm that netrin-1 activates endothelial cells through binding to CD146, we used the specific function-blocking CD146 antibody, AA98. Interestingly, AA98 could block both netrin-CD146 and CD146-VEGFR2 interactions [34], thus serving as a bi-functional blocker. As shown in Supplementary information, Figure S3, netrin-1 promoted HUVEC proliferation, migration, tube formation and downstream signaling activation in the control mIgG-treated groups. However, pre-treatment of HUVECs with AA98 abolished the effect of netrin-1 (Supplementary information, Figure S3), supporting that the netrin-CD146 and CD146-VEGFR2 interactions are critical for netrin-1-induced endothelial cell activation.

CD146 is required for netrin-1-induced angiogenesis in vivo

We further examined the role of netrin-CD146 interaction in two angiogenesis models. In the aortic-ring assay, endothelial cells sprout from the cultured mouse

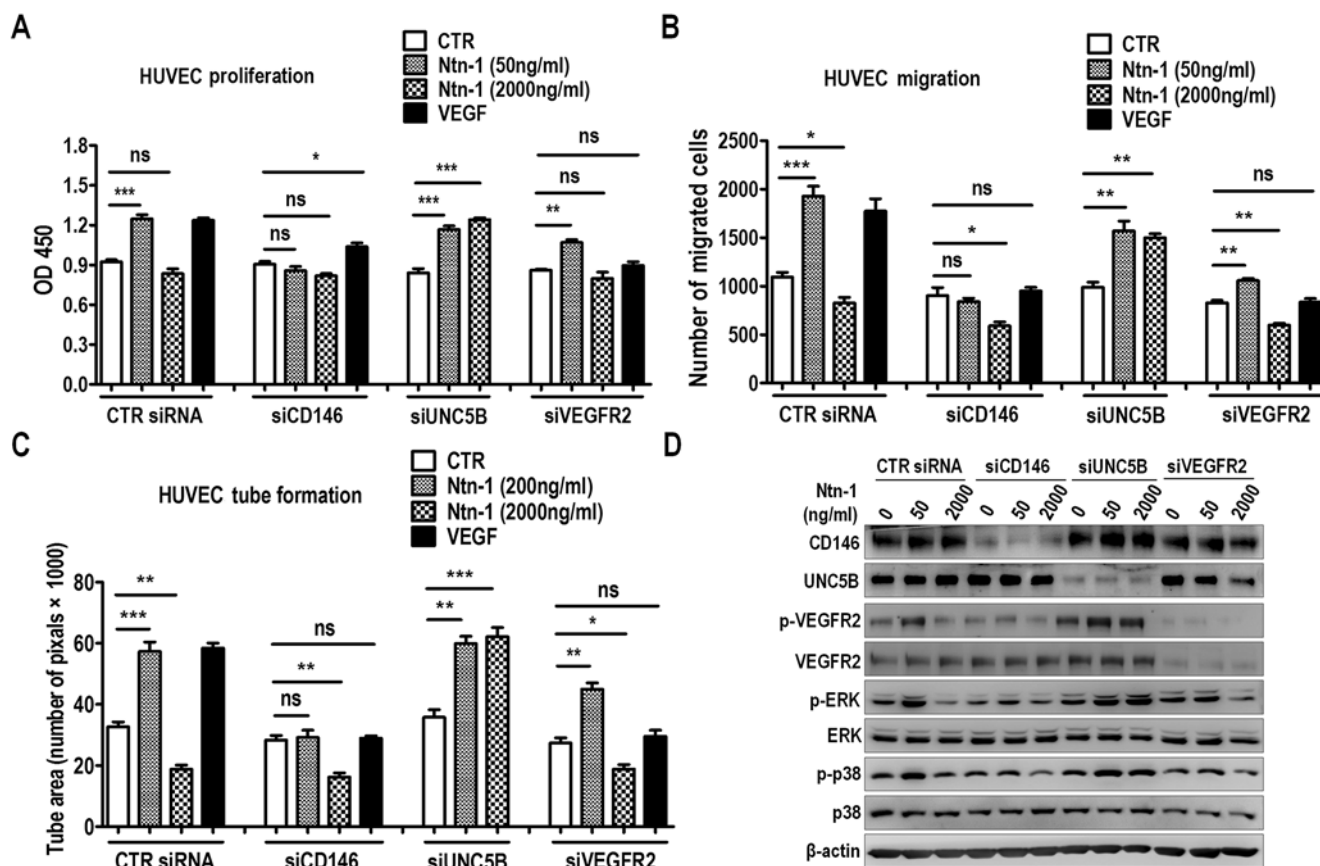


Figure 4 CD146 mediates netrin-1-induced endothelial cell activation. (A–D) HUVECs transfected with control, CD146, UNC5B or VEGFR2 specific siRNA were subjected to proliferation assay (A), transwell migration assay (B), tube formation assay (C) and signaling activation assay (D). Netrin-1 was applied at the indicated concentrations. Note that the specific siRNAs efficiently downregulated the expression of corresponding molecules. For A–C, $n = 3$ in each group. Data represent 3 independent experiments (means \pm SEM). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; 'NS' indicates no significant changes observed (one-way ANOVA with Turkey *post hoc* tests).

aortic explants and form tubular structures. In the Matrigel-plug assay, mouse endothelial cells migrate into the Matrigel implants where they form networks resembling capillaries. We first tested the effect of netrin-1 at different concentrations on these processes. Netrin-1 promoted endothelial cell sprouting and angiogenesis at 50 or 200 ng/ml, whereas it showed no or inhibitory effect at a higher concentration of 1 000 ng/ml (Supplementary information, Figure S4A and S4B), consistent with the results described in the endothelial cell activation.

Importantly, the average number of sprouts per ring was increased by netrin-1 (50 ng/ml) treatment of aortic rings from the wild-type (WT) mice (Figure 5A). However, when the aortic rings were prepared from CD146^{EC-KO} mice in which CD146 was conditionally deleted in the endothelium [39], the promoting effect of netrin-1 on endothelial cell sprouting was eliminated (Figure 5A). Consistently, anti-CD146 mAb AA98 blocked the stimu-

lating effect of netrin-1 on endothelial sprouting (Figure 5B).

In the Matrigel-plug assay *in vivo*, netrin-1 (200 ng/ml) increased the number of blood vessels in the Matrigel implants in the WT mice (Figure 5C). However, when CD146^{EC-KO} mice were used (Figure 5C) or when the Matrigel plugs were mixed with AA98 before injection (Figure 5D), the stimulatory effect of netrin-1 on vascular formation was significantly reduced, but not totally abolished. These results suggest that netrin-1-induced angiogenic response *in vivo* mainly depends on its binding to the endogenous endothelial CD146, although these data do not exclude the possibility that other mechanisms might also be involved.

To investigate the role of netrin-1 and CD146 in angiogenesis during vertebrate development, we used a zebrafish model. *In situ* hybridization revealed strong CD146 expression on endothelial cells at 24–36 hours

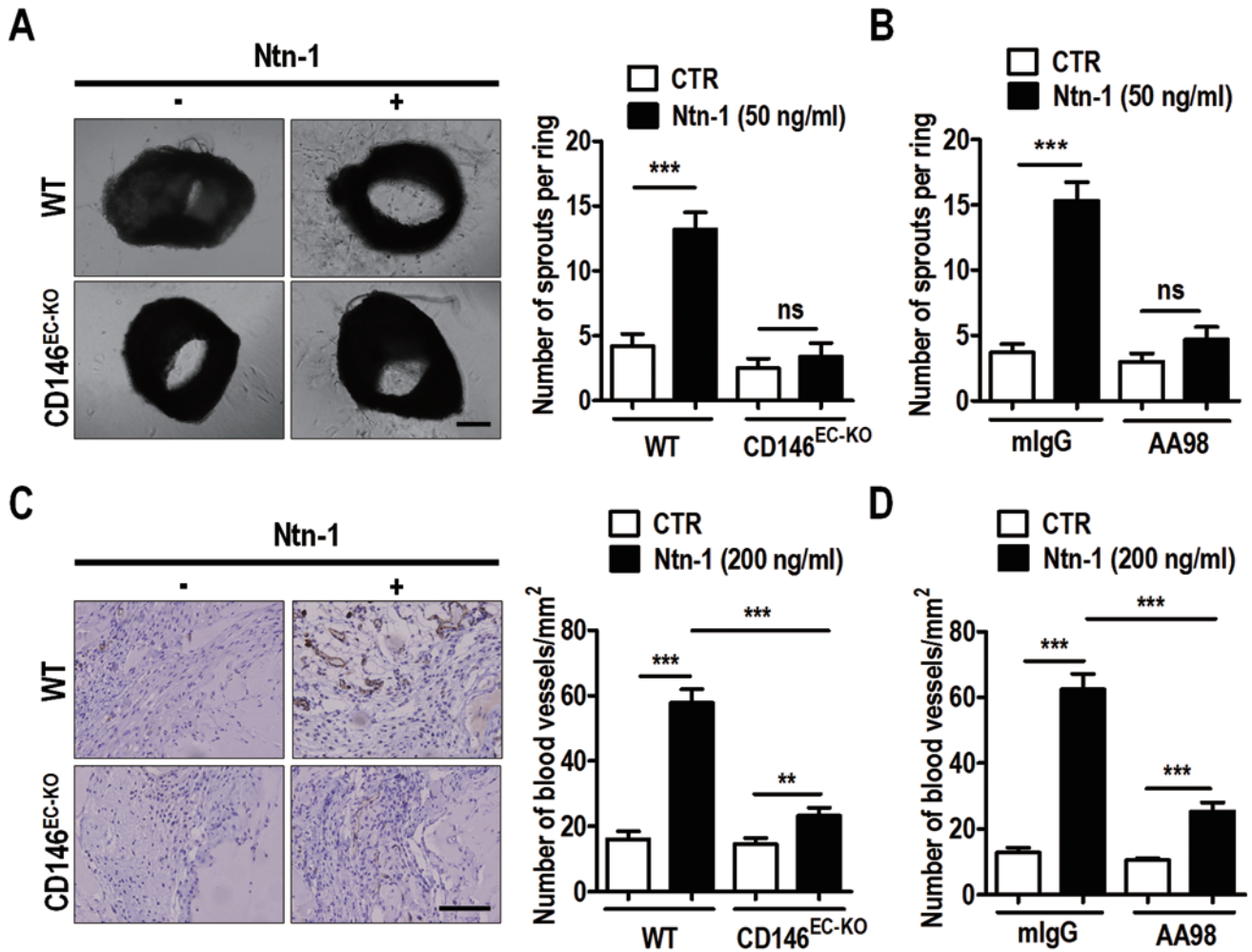


Figure 5 CD146 is required for netrin-1-induced angiogenesis in mouse models. **(A)** Aortic rings were prepared from WT or CD146^{EC-KO} mice. Control or netrin-1 (50 ng/ml) was directly added to the culture medium. **(B)** The effect of anti-CD146 antibody AA98 was tested in the aortic-ring assay. Control mlgG or AA98 (100 µg/ml) was added to the culture medium in the presence of control or netrin-1 (50 ng/ml). After culturing for 5-6 days, the number of sprouts from each ring was quantified. $n = 10$ in each group and results are presented as average number of sprouts per ring (means \pm SEM). **(C)** The Matrigel-plug assay for angiogenesis was carried out using WT or CD146^{EC-KO} mice. The plugs were mixed with control or netrin-1 (200 ng/ml) and then injected subcutaneously into mice in the corresponding groups. **(D)** The effect of anti-CD146 antibody AA98 on netrin-1-induced angiogenesis was tested in the Matrigel-plug assay. The plugs were pre-mixed with AA98 or control mlgG (100 µg/ml) and injected into the WT mice. 10 days post injection, the Matrigel plugs were sectioned and immunostained with anti-CD31 antibody. The number of blood vessels in each section was scored. $n = 5$ in each group and results are presented as average number of blood vessels/mm² (means \pm SEM). Scale bar, 200 µm. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns, not significant.

post fertilization (hpf), the critical stage for vascular development (Figure 6A). After 48 hpf, the CD146 expression was downregulated. This developmental expression pattern of CD146 on endothelial cells is consistent with its role in vascular development. Next, we used specific antisense morpholino oligonucleotides (MOs) to examine the functions of netrin-1a and CD146 in angiogenesis and vascular development. The corresponding

MOs efficiently knocked down the expression of the fish netrin-1a or CD146 (Supplementary information, Figure S5). In the transgenic Tg(kdrl:GFP) embryos which express GFP in the endothelial cells to enable visualization of the vasculature, downregulating the zebrafish CD146 homolog by the CD146 MO led to a phenotype that was strikingly similar to that of the netrin-1a MO-treated embryos: the intersegmental vessels (ISVs) and dorsal

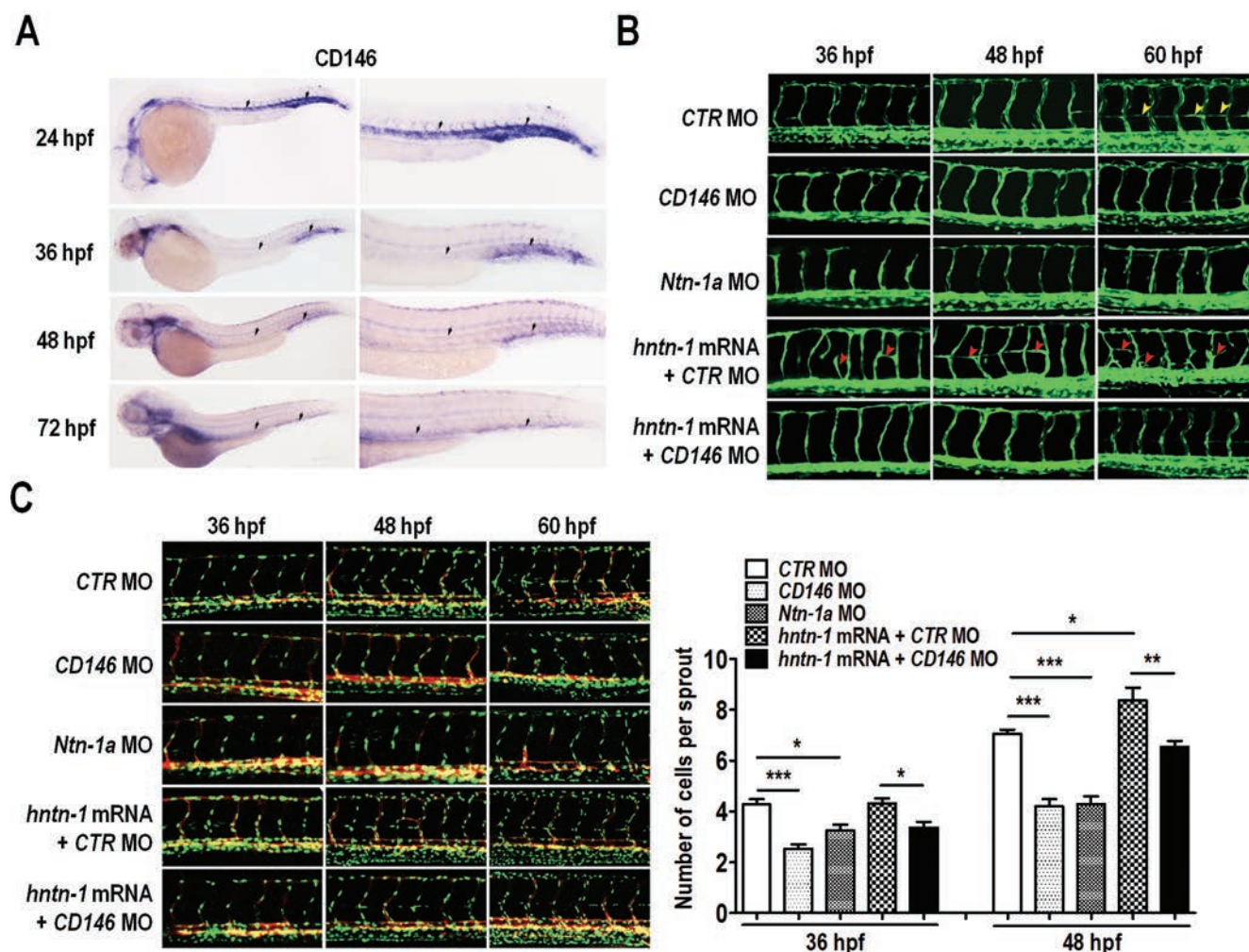


Figure 6 CD146 mediates netrin-1-induced angiogenesis in zebrafish. **(A)** CD146 expression in the zebrafish embryos at different stages was detected by whole mount *in situ* hybridization. **(B)** The control, *CD146* or *netrin-1a* specific MO was injected with control or human netrin-1 mRNA into the transgenic zebrafish Tg(kdrl:GFP) embryos that expressed GFP to enable visualization of vasculature. The vascular patterns of the fish embryos at 36, 48 and 60 hpf were analyzed. Yellow arrowheads indicate PAV. Red arrowheads mark ectopic sprouts in netrin-1 mRNA-injected embryos. **(C)** Tg(Fli1:nGFP) × Tg(kdrl:mcherry) embryos were injected with the indicated MO and mRNA. The number of endothelial cells in the ISVs of the embryos at 36 hpf, 48 hpf and 60 hpf stages was quantified and analyzed. Data are presented as average number of endothelial cells per ISV (means ± SEM). **P* < 0.05; ***P* < 0.01; ****P* < 0.001 (one-way ANOVA with Turkey *post hoc* tests).

longitudinal anastomotic vessels (DLAVs) were formed, whereas the formation of parachodal vessels (PAVs) was inhibited (Figure 6B). *CD146* morphants (38/45) and *netrin-1a* morphants (30/39) lacked *fli:egfp* positive cells at the presumptive position of PAV. Furthermore, overexpressing human netrin-1 (*hntn-1*) in the fish embryos caused ectopic vessel sprouting and branching (36/43), which was blocked by co-injection of *CD146* MO (41/41). These data demonstrate that netrin-1-induced angiogenesis depends on *CD146* expression during zebrafish vascular development.

We further tested the effect of netrin-1 and *CD146*

on endothelial cell proliferation and migration using *Fli1:nGFP* embryos. At the stages of 36, 48 and 60 hpf during vascular development, *CD146* MO or *netrin-1a* MO injected embryos showed a reduction of endothelial cells in the ISVs (Figure 6C). Time-lapse microscopy showed that both proliferation and migration of endothelial cells were inhibited by *netrin-1a* MO or *CD146* MO injection (Supplementary information, Figure S6). Consistently, increased endothelial cells in the ISVs were observed in embryos overexpressing *hntn-1*, and *CD146* MO blocked the increase of endothelial cells caused by *hntn-1* overexpression (Figure 6C). Taken together,

these results demonstrate that CD146 and netrin-1a are crucial for vascular development in zebrafish, and netrin-1 induces endothelial cell activation and angiogenesis in a CD146-dependent manner.

Discussion

The role of netrin in angiogenesis is highly complex. By interacting with different receptors, netrin can act as a pro- or anti-angiogenic factor. In the present study, we demonstrate that CD146 mediates netrin-1-promoted endothelial cell activation and angiogenesis *in vitro* and *in vivo* with several lines of evidence. First, netrin-1 specifically binds to CD146 *in vitro* with high affinity in the absence of other proteins. Second, siRNA-mediated knockdown of CD146 or inhibition of netrin-CD146 interaction by the anti-CD146 mAb AA98 suppresses netrin-1-induced endothelial cell proliferation, migration and tube formation, as well as the activation of downstream signals. Third, endothelial-specific CD146 deletion or the treatment with the CD146-specific mAb AA98 inhibits netrin-1-promoted endothelial cell sprouting and blood vessel formation in mice. Fourth, down-regulation of netrin-1a or CD146 by specific MO results in similar defects in zebrafish vascular development. CD146 knockdown eliminated ectopic vascular sprouting induced by netrin-1 overexpression. Collectively, our findings demonstrate the ligand-receptor relationship between netrin-1 and CD146, and support a critical role of netrin-CD146 signaling in angiogenesis during vertebrate vascular development.

Previous studies have demonstrated the dual role of netrin-1 in angiogenesis, and the pro- or anti-angiogenic function of netrin-1 depends on its concentration [4, 37, 40]. At high concentrations, netrin-1 inhibits angiogenesis through UNC5B receptor [7, 21]. Here, we demonstrate netrin-1 at relatively low concentrations promotes angiogenesis through CD146. Netrin-1 binds to CD146 with a higher affinity than that of netrin-UNC5B interaction, as shown by the SPR experiments. Moreover, in the endothelial cell line, CD146 is expressed at a much higher level than UNC5B at both RNA and protein levels (Supplementary information, Figure S7A and S7B). We propose that one explanation for the dual role of netrin-1 is the different expression levels of CD146 and UNC5B receptors. At low concentrations, netrin-1 preferentially binds to CD146, whereas at high concentrations, netrin-1 could bind to UNC5B and triggers signals that counteract CD146 downstream signals to inhibit the growth of blood vessels. It will be interesting to systematically compare the signal pathways downstream of netrin-1 receptors involved in pro- and anti-angiogenesis processes,

and to exam how UNC5B and CD146 coordinate with each other to mediate netrin-1 signaling in directing morphogenesis of the vascular system during development. Further studies are necessary to delineate these pathways and elucidate molecular mechanisms underlying these processes.

In our Matrigel-plug model, the effect of netrin-1 on angiogenesis was substantially diminished, but not fully eliminated by endothelial CD146 knockout or AA98 treatment. It is conceivable that the involvement of multiple factors in the *in vivo* microenvironment may have contributed to the results. Although endothelial CD146 is deleted in CD146^{EC-KO} mice, there are other types of cells expressing CD146 which might be activated by netrin-1 to promote angiogenesis. The smooth muscle cells and macrophages express CD146 and are tightly connected to the progression of angiogenesis. Moreover, other netrin receptors might be expressed on various types of cells and contribute to netrin-1-induced angiogenesis *in vivo*. Therefore, activating endothelial cells through CD146 is one important pathway via which netrin-1 promotes angiogenesis, and this mechanism may operate in conjunction with other mechanisms *in vivo*.

Using zebrafish models, we demonstrate that knockdown of netrin-1a or CD146 causes specific vascular defects, supporting the notion that netrin-CD146 plays a critical role in vascular development. In addition to its role in developmental angiogenesis, netrin-CD146 signaling might be involved in pathological angiogenesis. Expression of netrin-1 in melanoma, breast cancer and colorectal cancer confers metastatic advantages to tumor cells [41-43], supporting a role of netrin-1 in tumor angiogenesis. The involvement of endothelial CD146 in tumor angiogenesis has been extensively documented. Based on these findings and the present work, we propose that netrin-CD146 signaling may also have a role in diseases involving pathological angiogenesis. Modulating netrin-CD146 signaling by specific antibodies or analogues may provide new therapeutic approaches to angiogenic diseases.

In our previous study, we have demonstrated that CD146 promotes VEGF-induced tumor angiogenesis by acting as a co-receptor for VEGFR2. However, the CD146-knockout mice did not show severe vascular defects and embryonic lethal phenotype [39], which were observed in VEGF-knockout mice [44, 45] and VEGFR2-knockout mice [46]. These data suggest that under physiological conditions, CD146 and VEGF signaling play different roles in regulating angiogenesis. Indeed, our present study demonstrates that CD146 is a receptor for netrin-1 to promote developmental angiogenesis. Moreover, CD146 acts as a receptor for Wnt5a to regu-

late cell motility and convergent extension [26], and Wnt signaling has been reported to contribute to angiogenesis as well [47, 48]. The schematic of CD146 involved, angiogenesis-related pathways is shown in Supplementary information, Figure S8. It seems that CD146 can mediate different signalings according to the context, and further studies should be performed to demonstrate how these pathways crosstalk and cooperate with each other.

Material and Methods

Antibodies and reagents

The following antibodies were used in this study: anti-CD146 rabbit polyclonal antibody, anti-CD146 mouse mAb AA1 and AA98 were generated in our lab [29, 36]; the control mIgG, anti- β -actin and anti-Flag antibodies (Sigma-Aldrich); anti-netrin-1 and anti-UNC5B antibodies (Enzo Life Science); anti-p38, anti-phosphor-p38, anti-ERK1/2 and anti-phosphor-ERK1/2 antibodies (Cell Signaling Technology); anti-VEGFR2 and anti-phosphor-VEGFR2 antibodies (Signalway antibody); anti-CD31 antibody (Abcam); anti-human IgG Fc and anti-His antibodies (ZSGB-BIO); HRP-conjugated goat anti-mouse and -rabbit IgG antibodies (GE Healthcare); goat anti-mouse Alexa Fluor 555 (Invitrogen); biotin-conjugated goat anti-rabbit IgG antibody and HRP-conjugated streptavidin (Dianova).

The following reagents were used: recombinant hnetrin-1, mouse netrin-1 and Fc-UNC5B (Enzo Life Science); human Fc, Fc-CD146, sCD146 (Sino Biological), growth factor-reduced Matrigel (BD Biosciences); Fugene HD, DAPI and protease inhibitor cocktails (Roche); protein G sepharose beads and DAB substrates (Santa Cruz); Enhanced Chemiluminescence Assay Kit (Pierce) for western blotting; cell counting kit-8 (CCK8, Dojindo) for cell proliferation assay.

Animals

All animal experiments were performed in compliance with national guidelines for the care and use of laboratory animals and were approved by the Biomedical Research Ethics Committee of the Institute of Biophysics, Chinese Academy of Sciences. Tek^{cre/+}CD146^{flxed/flxed} (named CD146^{EC-KO}) mice were generated in our lab as described previously [39]. Tek^{cre/+}CD146^{flxed/flxed} (WT) mice were used as controls in the aortic-ring and the Matrigel-plug assays.

Zebrafish embryos were obtained by natural spawning of adult fish. Embryos were raised and maintained at 28.5 °C and staged morphologically as described previously [49]. The following transgenic lines were used: Tg(fli1:nEGFP), Tg(kdrl:EGFP) and Tg(kdrl:mCherry).

Co-immunoprecipitation

HEK293 cells were co-transfected with plasmids encoding the full-length CD146, netrin-1 or the truncation mutants using Fugene HD (Roche). 48 h post transfection, the cells were lysed in ice-cold RIPA buffer (150 mM NaCl, 50 mM Tris, pH 8.0, 0.1% SDS, 0.5% deoxycholate, 0.1% NP-40, 1 mM PMSF, protease inhibitor cocktails). Then the cell lysates were incubated with the control mIgG, anti-CD146 mAb AA1 or anti-netrin-1 mAb for 4 h at 4 °C, and immunoprecipitation was carried out using protein G sepharose beads (Santa Cruz). The cell lysates and precipitates were analyzed

by western blotting using chemiluminescence imaging system (ChemiScope 3400; Clinx, China).

Pull-down assay

For pull-down assay, purified human Fc, Fc-CD146 or Fc-UNC5B (200 ng/ml each) was immobilized on protein G sepharose beads. Beads were then incubated with recombinant hnetrin-1 (200 ng/ml) for 2 h at 4 °C in HEPES buffer (0.05 M HEPES, 0.15 M NaCl, 0.001% Tween 20, pH 7.4). After three washes in HEPES buffer, the bound proteins were analyzed by western blotting.

SPR assay

Fc-CD146, Fc-UNC5B or Fc (10 μ g/ml each) in HEPES buffer was immobilized on the surface of a CM5 sensor chip (BIAcore AB) docked into a BIAcore 3000 instrument (flow rate: 5 μ l/min) using the amine coupling kit (BIAcore AB) following the manufacturer's instructions. Purified netrin-1 at different concentrations (18.75 nM, 37.5 nM, 75 nM, 150 nM and 300 nM) was injected at a flow rate of 5 μ l/min and the response was measured in RU (Resonance Units). A blank flow cell was prepared by injecting netrin-1 with the same flow rate over a blank sensor chip to obtain the baseline RU. The net RU reported was obtained by subtracting the baseline RU from the response RU. The Fc protein was used as a negative control.

RNA interference

CD146- and UNC5B-specific siRNAs were synthesized by Invitrogen using sequences as previously described [27, 37]. Downregulation of VEGFR2 in HUVECs was carried out using a shRNA kit (Origene). HUVECs were transfected using Fugene HD (Roche) and functional or signaling assays were carried out 48 h post transfection.

Endothelial cell proliferation, migration and tube formation

In proliferation, migration and tube formation assays, HUVECs that had been serum-starved for 24 h were used. Cell proliferation assay was carried out using a CCK-8 kit (Dojindo). Briefly, HUVECs were suspended and seeded into a 96-well plate. Netrin-1 and the antibodies were directly added to the culture medium. After culturing for 48 h, cells were incubated with 100 μ l of 10% CCK-8 reagent for 1 h at 37 °C. The color reaction was measured at 450 nm with a BioRad ELISA reader (Richmond, CA, USA).

HUVEC migration was examined using a modified Boyden chamber assay (8 μ m pore size; Costar, Corning, USA). HUVECs were suspended in fresh serum-free medium and seeded in upper chambers of the transwell plate to a total number of 5×10^3 cells per chamber. Lower chambers contained fresh medium containing 10% fetal calf serum (FCS). After 12-h incubation at 37 °C, cells remaining at the upper surface of the membrane were removed using a swab, whereas the cells migrated to the lower membrane surface were fixed with 4% PFA and stained with 0.1% crystal violet solution. The number of cells migrating through the filter was scored.

In tube formation assay, 96-well culture plates were coated with Matrigel (BD Biosciences) to a total volume of 60 μ l per well and allowed to solidify for 30 min at 37 °C. Netrin-1 was added directly to the HUVEC suspensions in complete 1640 medium, and the cells were then seeded into the corresponding wells to a total number of 1×10^4 cells per well in 96-well plate. Cells were incubated at 37 °C for 6 h. Tube formation was observed under an inverted

microscope (Eclipse Model TS100; Nikon, Japan) and tube length was measured using NIH Image J software.

Signaling activation assays

HUVECs were serum-starved for 24 h and then treated with netrin-1 at different concentrations for 10 min at 37 °C. The cells were lysed in ice-cold RIPA buffer and boiled in sample loading buffer before western blotting analysis. For antibody blocking test, HUVECs were incubated with CD146-specific mAb AA98 or control mIgG (50 g/ml) for 1 h before netrin-1 was added.

The aortic-ring assay

The aortic-ring assay was performed as described [50]. Briefly, thoracic aortae were carefully dissected from 12-week-old mice following euthanasia and cut into rings of 0.5-1 mm in width. The rings were then serum-starved overnight in Opti-MEM medium and implanted into Matrigel in 96-well culture plate. The rings were cultured with Opti-MEM containing 2.5% FCS for 5-6 days (growth medium was changed every other day). Then the rings were photographed under an inverted microscope, and the number of sprouts per ring was quantified. For the antibody blocking experiment, mIgG or AA98 (100 µg/ml) was added directly to the growth medium.

The Matrigel-plug assay

The Matrigel-plug assay was performed as described [34]. Liquid Matrigel containing heparin was mixed with control or netrin-1 (200 ng/ml). The Matrigel mixture (500 µl) was then injected into the abdominal subcutaneous tissue of mice along the peritoneal midline. Ten days post injection, the mice were sacrificed and the Matrigel plugs were carefully dissected out. After photographing, the plugs were fixed with 4% PFA in PBS and embedded into paraffin. Samples were sectioned at 5 µm and then immunostained with anti-CD31 antibody. The number of blood vessels in each section was quantified and analyzed. For antibody blocking assay, Matrigel was mixed with control mIgG or AA98 (100 µg/ml) before injection.

Immunohistochemistry

For immunohistochemistry staining, the paraffin-embedded Matrigel-plug sections were de-paraffinized and stained first with an antibody specific for endothelial marker CD31, followed by biotin-conjugated secondary antibody and HRP-conjugated streptavidin (Dianova). Sections were then counterstained with hematoxylin. The number of blood vessels in each section was scored under a microscope.

Morpholinos, mRNA synthesis and microinjection in zebrafish

The CD146-MO (5'-AGCAGTGC GGTGTAGGTCATTCTC-3') and control MO were purchased from GeneTools (Philomath, OR, USA) and prepared as 1 mM stock solutions. The netrin-1a-MO (5'-CCAAAGCATCAGAGACTCTCAACAT-3') was a gift from G Peng [51]. For CD146 or netrin-1a knockdown experiment, 2 ng CD146 MO or netrin-1a MO per embryo was injected at one-cell stage. The full-length hnetrin-1 cDNA was inserted into pCS2+ vector and mRNA for injection was synthesized *in vitro* using the mMessageMachine SP6 kit (Ambion) according to the instruction manual. For hnetrin-1 overexpression experiment, 12.5 pg *hnetrin1* mRNA per embryo was injected at one-cell stage. Confocal

images were acquired using a Zeiss LSM 510 META confocal microscope with 3D projections generated using Zeiss LSM software (Carl Zeiss) [52].

Statistical analysis

Unless specified otherwise, all data represent at least 3 independent experiments in each assay. Results are expressed as the mean ± SEM. One-way ANOVA with Turkey *post hoc* tests were used to compare differences between groups in various experiments. The criterion for statistical significance was defined as $P < 0.05$.

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